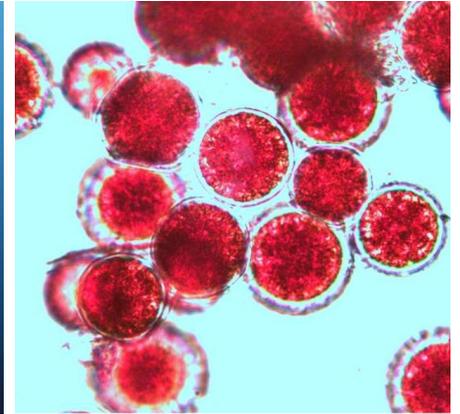
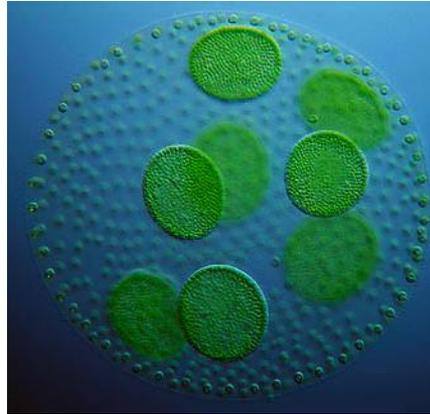
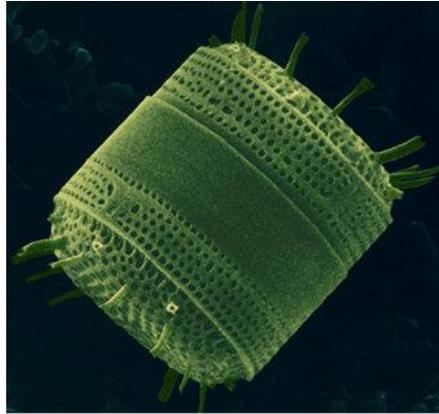
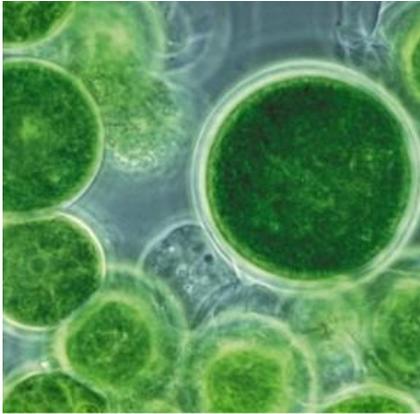


Advances in the development of genetic tools for microalgae research and biotechnology

BIO Pacific Rim Summit
2013

Beth A. Rasala, Ph.D.
Prof. Stephen Mayfield's Lab/
Triton Algae Innovations

Microalgae as a biomanufacturing platform



Biofuels

bio-diesel
bio-gasoline
jet fuel
bio-hydrogen
ethanol

Nutraceuticals

carotenoids
omega-3 fatty acids
astaxanthin
other vitamins and
supplements

Chemicals

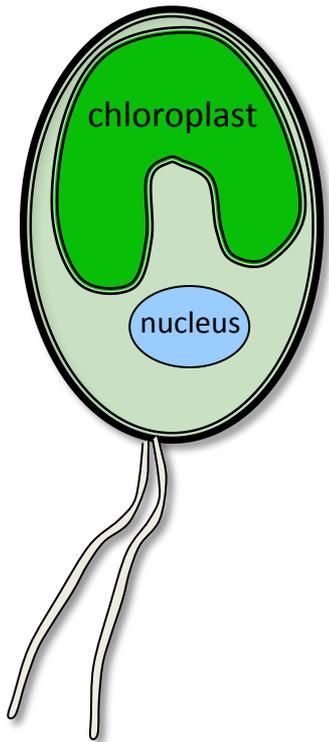
cosmetics
industrial
lubricants
bio-plastics
and more...

Proteins

therapeutics
oral vaccines
antimicrobials
industrial enzymes
animal feed

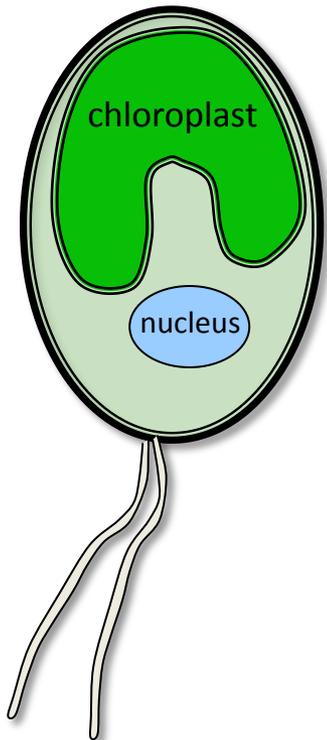
Genome engineering will likely enhance algal biotechnology.

Engineering *Chlamydomonas reinhardtii*



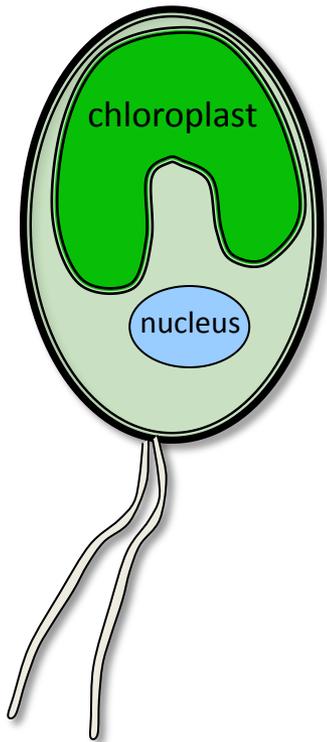
	Nucleus	Chloroplast
Transformation	Random integration	Homologous recombination
Stability of transgene	Susceptible to gene silencing	Highly stable once homoplasmic
Levels of heterologous protein accumulation	Low Levels susceptible to position effect	High Levels consistent across clones
Protein modification	Disulfide bond formation Glycosylation	Disulfide bond formation
Protein localization	Cytosol, organelle targeting, secretion	Protein retained in chloroplast

Chloroplast engineering – a success



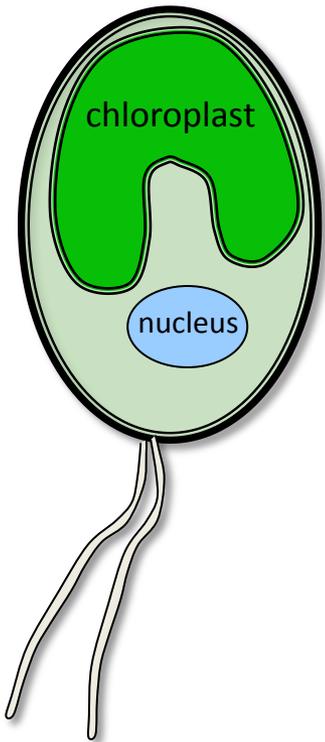
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Nuclear genome engineering



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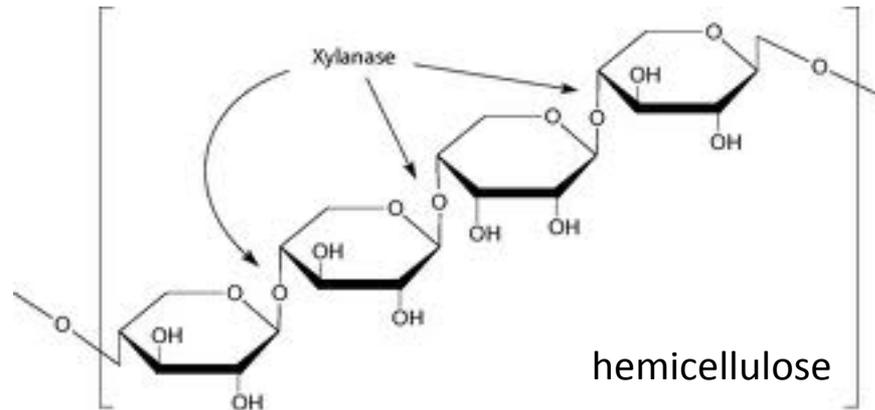
Nuclear genome engineering



	Nucleus	Chloroplast
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**Improved methods for
nuclear genome engineering
in green microalgae**

Case Study: Heterologous expression of Xylanase (*T. reesei*)



- **Endo-1,4-beta-xylanase (*Trichoderma reesei*)**
- hydrolysis of xylosidic linkages in hemicellulose polysaccharides
- industrial applications including paper manufacturing, **animal feed**, bread-making, juice and wine industries, and more
- potential large-scale use in **cellulosic ethanol**

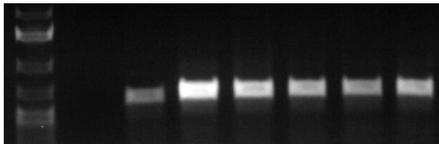
Heterologous expression of Xylanase



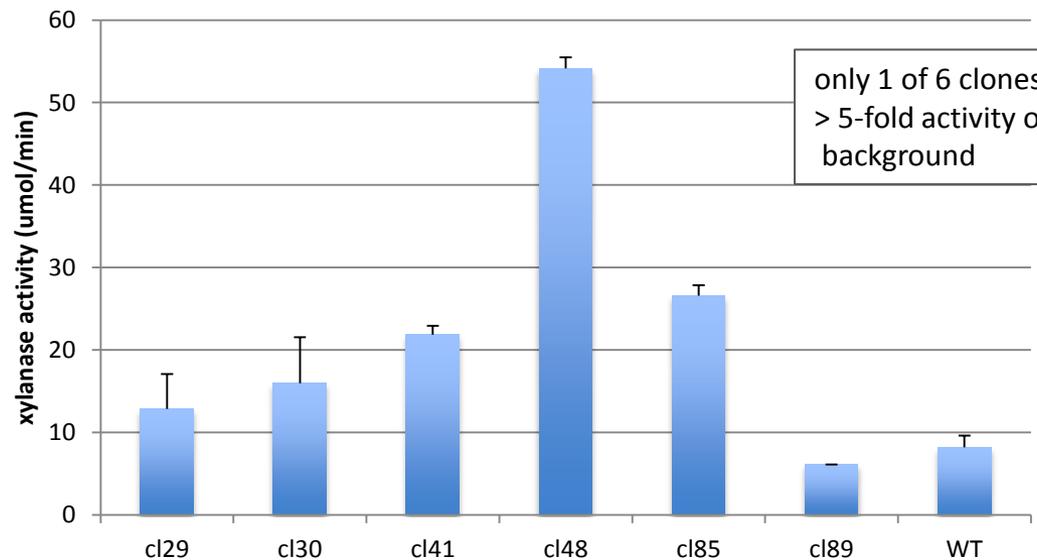
PCR: gene positive

P->xyn1

M WT cl29 cl30 cl41 cl48 cl85 cl89

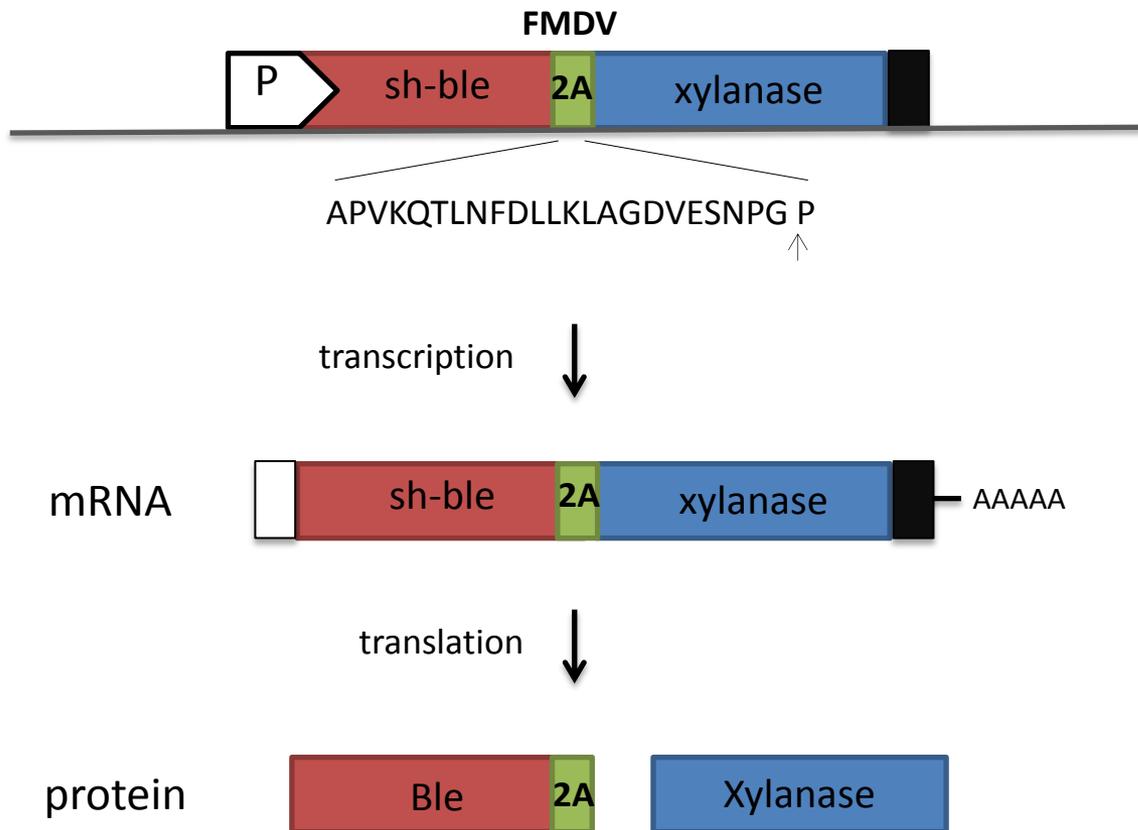


xylanase activity

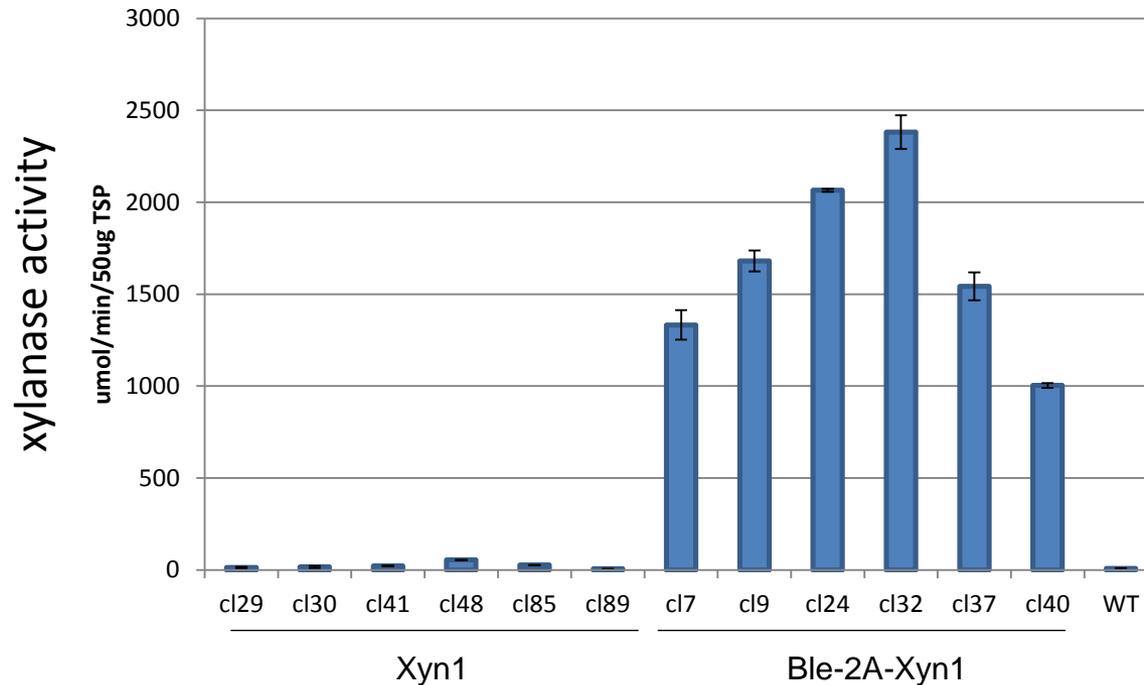
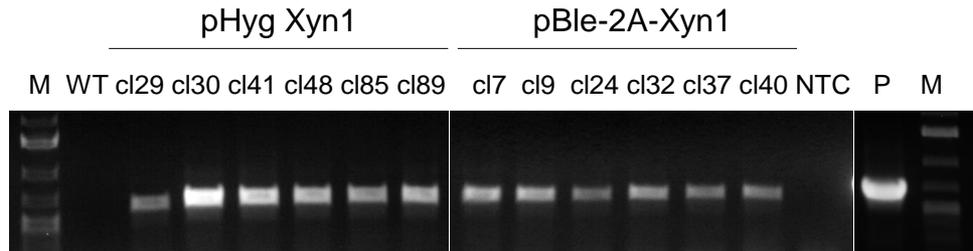


only 1 of 6 clones had > 5-fold activity over background

Transcriptionally linking *xylanase* expression to antibiotic resistance using the self-cleaving 2A sequence

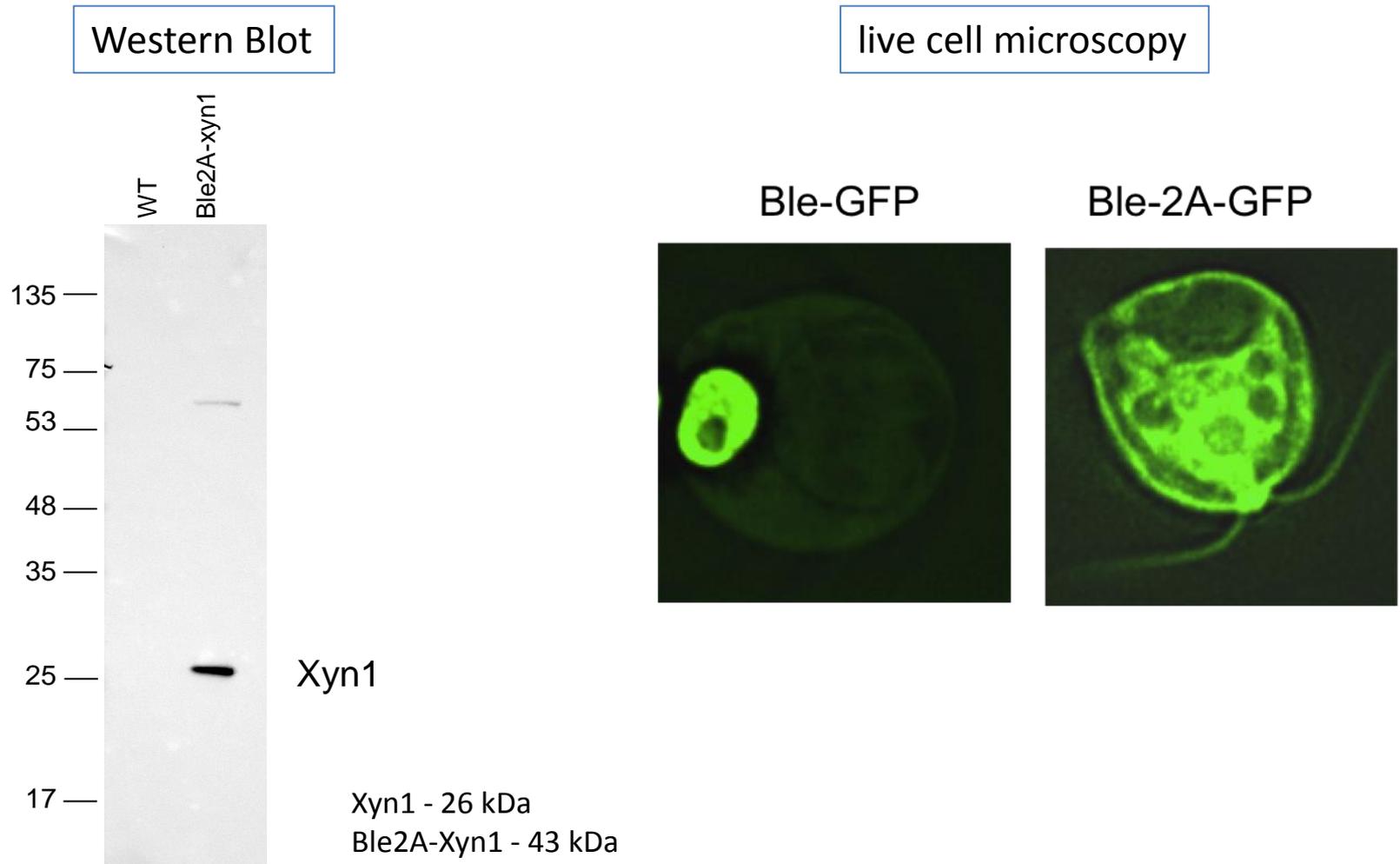


Transcriptionally fusing Xyn1 to Ble2A leads to the selection of transformants with higher xylanase activity

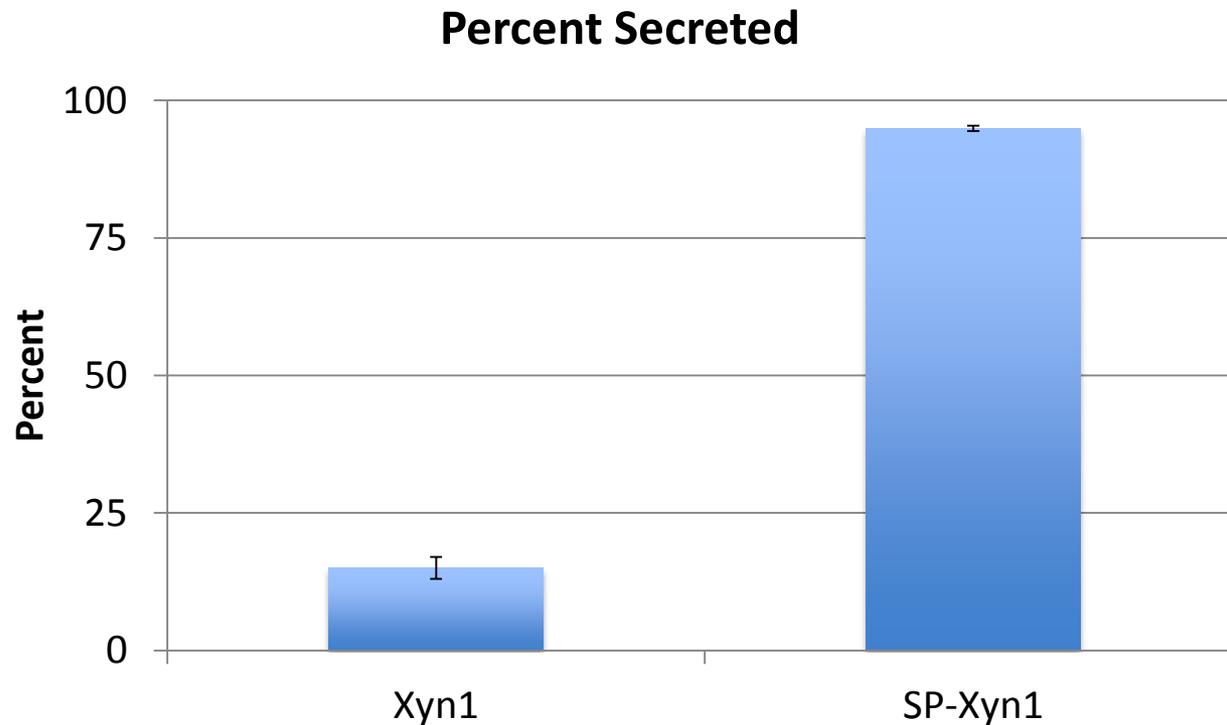


~ 100-fold
 higher activity

FMDV 2A sequence is 'cleaved' in microalgae



The Ble-2A vector supports xylanase secretion



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Chlamydomonas Protein Expression Kit

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Chlamydomonas Protein Expression Kit

[Synechococcus Protein Expression Kit](#)

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[Transformation Products for Algal Expression](#)

[GeneArt® Cryopreservation Kit for Algae](#)



The GeneArt® *Chlamydomonas* Protein Expression Kit is a second-generation *Chlamydomonas* cloning and expression system from Life Technologies. Like the first-generation GeneArt® *Chlamydomonas* Engineering Kit, this new system offers transgene expression from the nuclear genome of eukaryotic green alga *Chlamydomonas reinhardtii* 137c, but is optimized for high-level expression, provides selection against gene silencing, and offers dual protein tags for detection and/or purification of your gene of interest. It includes frozen *Chlamydomonas reinhardtii* 137c cells, MAX Efficiency® Transformation Reagent for Algae, expression vector, OneShot® TOP10 Competent *E. coli* cells, and easy-to-follow protocols. Our Gibco® TAP Growth Media, offered separately, is optimized for the growth and maintenance of *Chlamydomonas*.

- Express up to 1% total soluble protein of your gene of interest
- Select against gene silencing, even over multiple passages
- Detect and purify your gene of interest with 6His TEV and/or V5-His epitope tags
- Use (optional) [seamless assembly](#) to create your constructs
- Enables reliable results with exceptional strain viability and purity

Enabling fluorescent protein tools and technologies

UC San Diego

life
technologies™

UNIVERSITY OF
Nebraska
Lincoln

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UNIVERSITY

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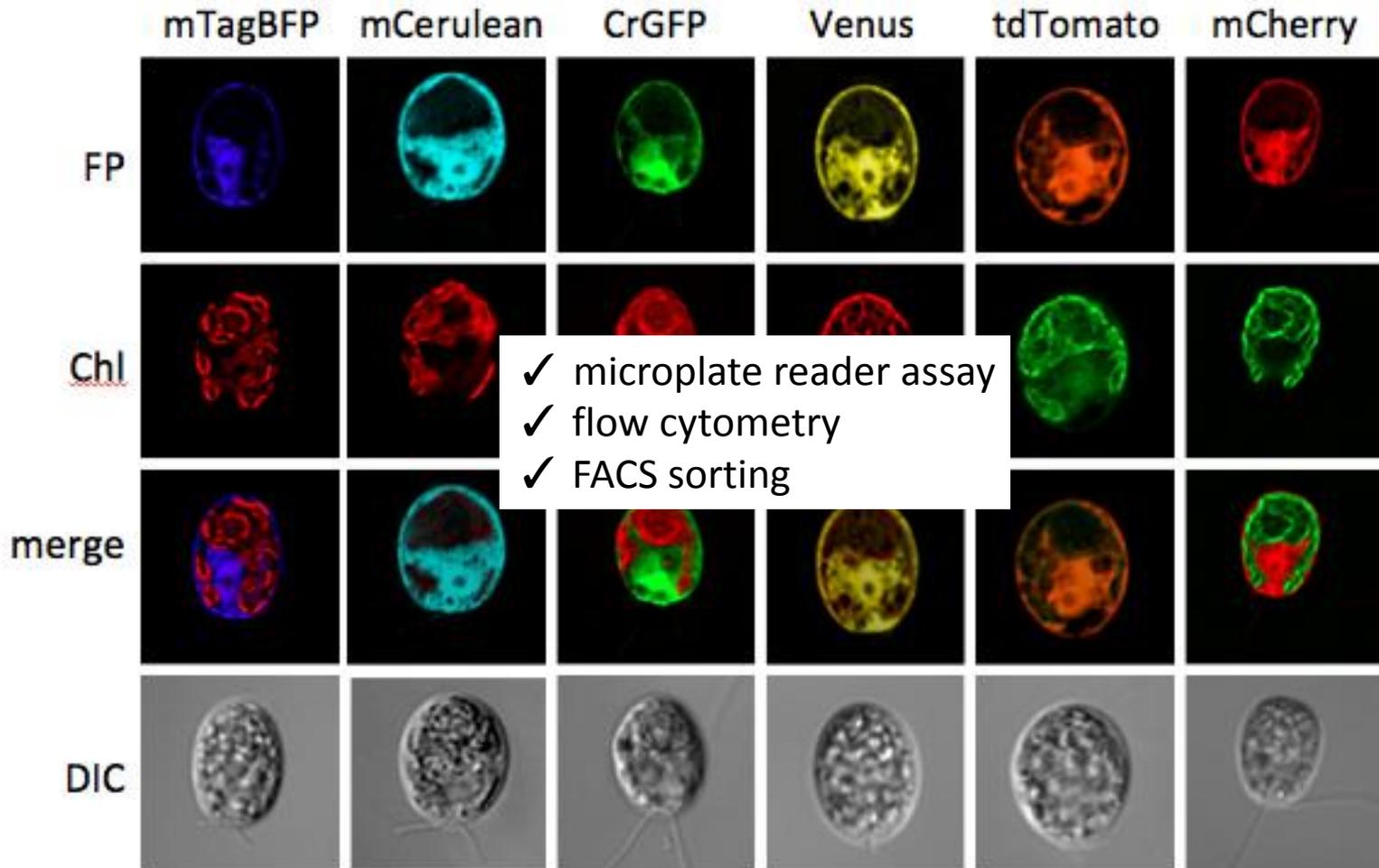
Development of **fluorescent reporters** in microalgae

Fluorescent protein (FP) technology has revolutionized many fields of biology, with an ever-expanding list of applications, including the investigation of:

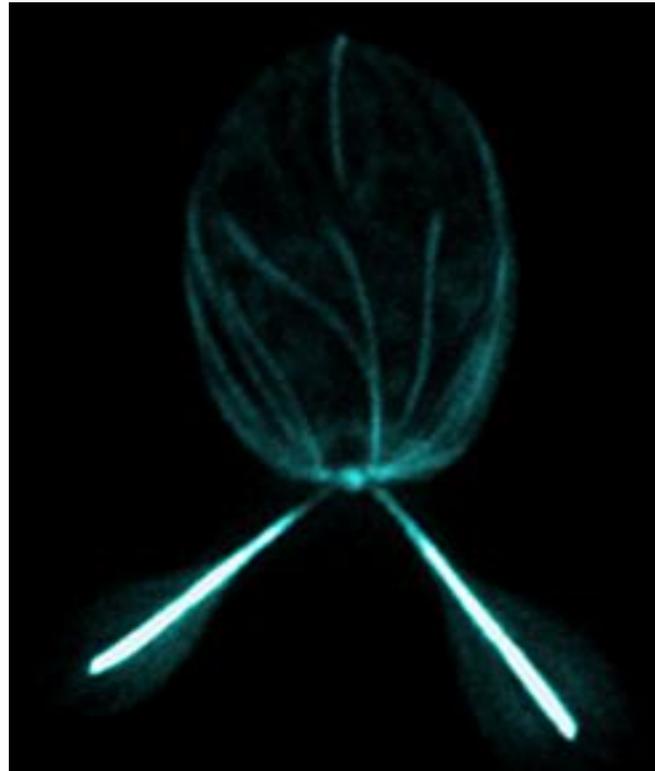
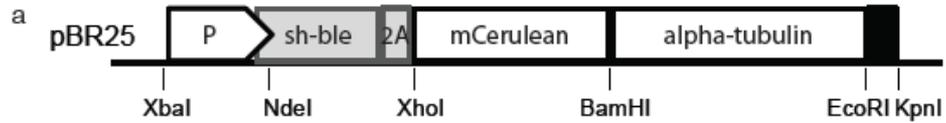
- promoter function
- protein localization
- protein-protein interactions
- cell cycle progression
- organelle labeling
- and as biosensors for monitoring small molecules and second messengers.

However, due to poor transgene gene expression from the nuclear genome, FPs have not been widely developed for microalgal research.

Live Cell Microscopy: FP expression in *C. reinhardtii*



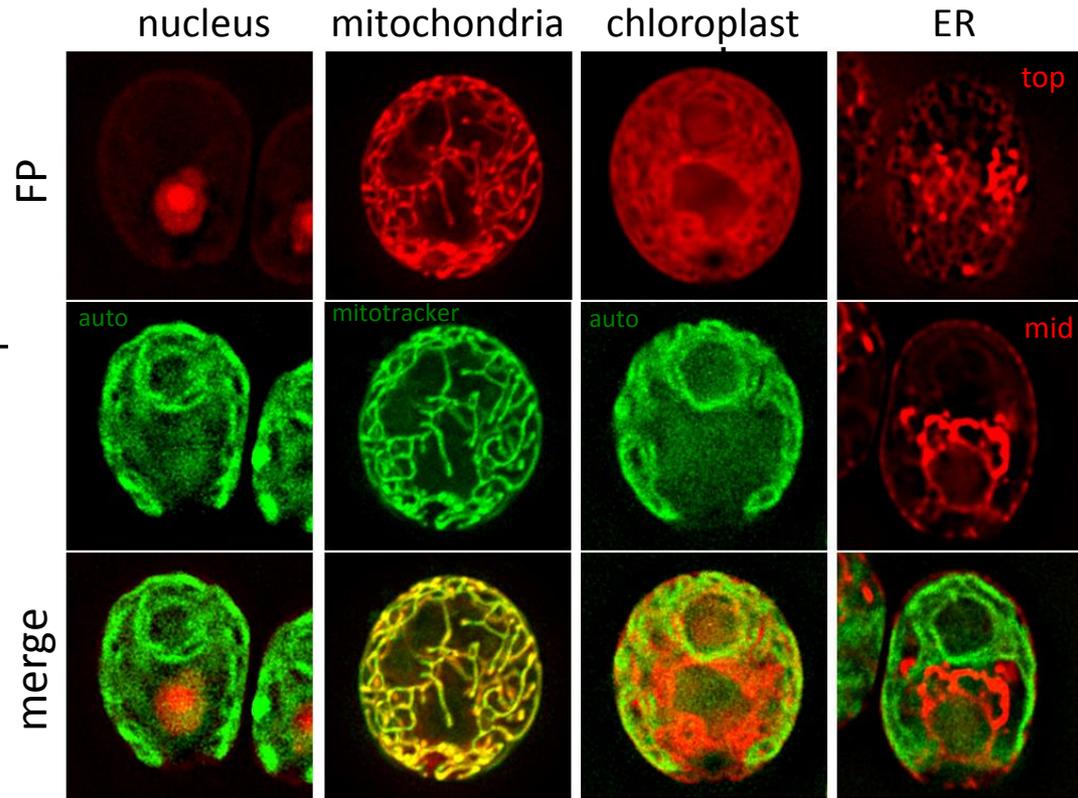
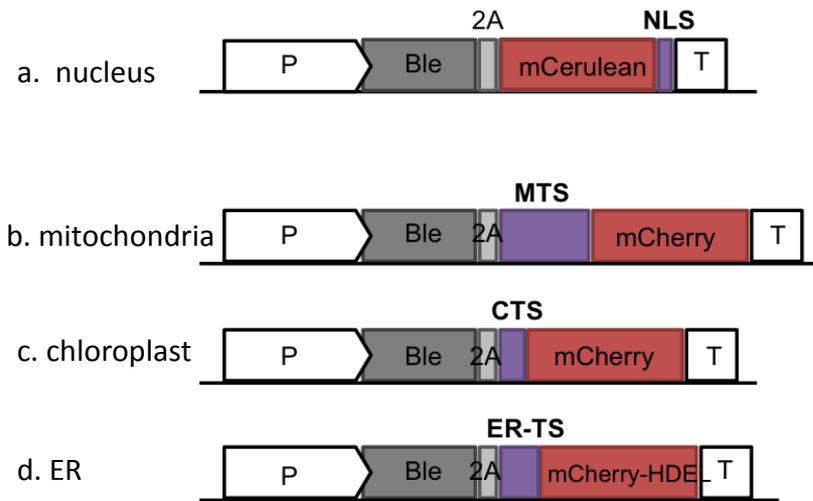
Expression of a mCerulean- α -tubulin fusion protein in *Chlamydomonas*



Enabling metabolic engineering

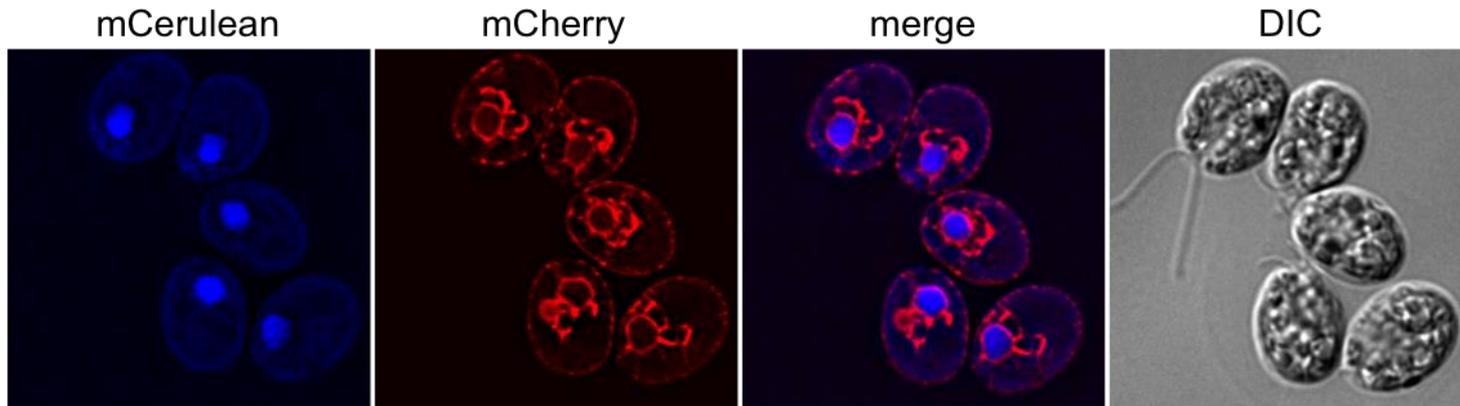
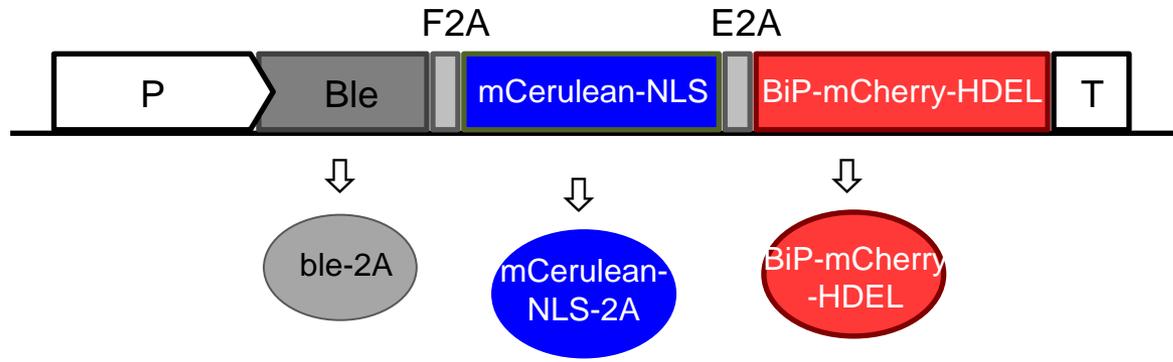
- Subcellular targeting vectors
- Coordinated expression of multiple transgenes within a single cell

Organelle targeting vectors for advanced genetic engineering



live cell microscopy

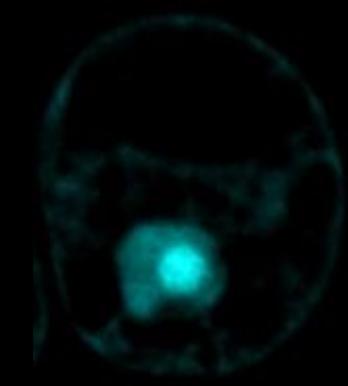
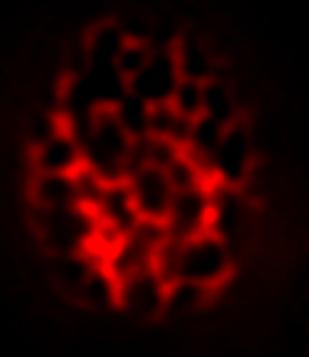
Gene stacking using a single transformation vector



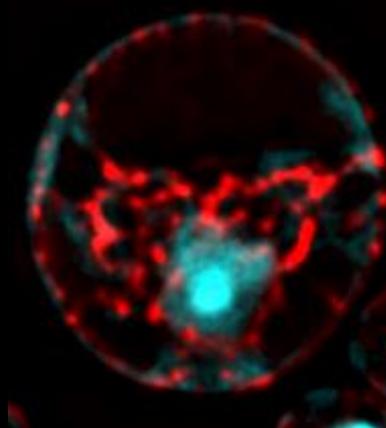
Gene stacking through mating

mCherry -> ER

mCerulean -> nucleus

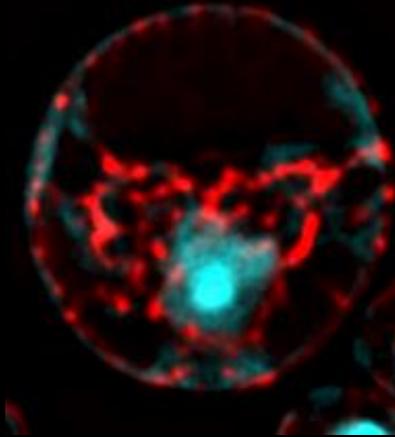


1st
X

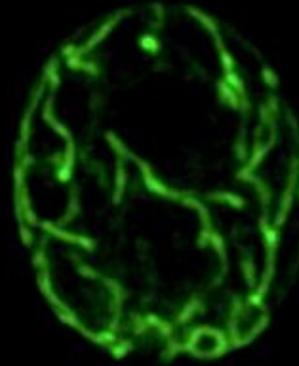


Gene stacking through mating

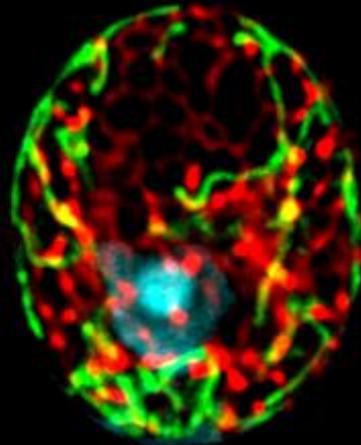
ER/nucleus



Venus -> MT

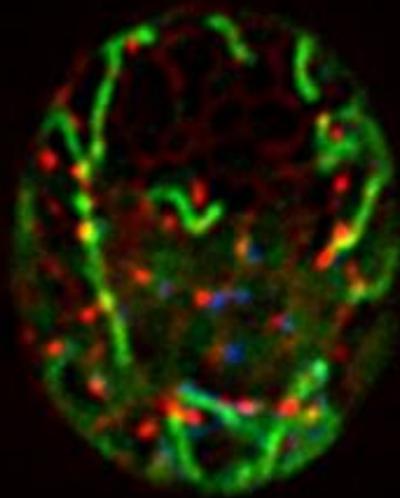


2nd
X

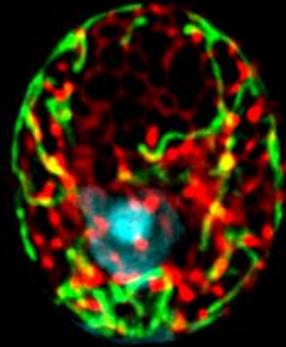


A Z-stack through *C. reinhardtii*

ER
nucleus
mitochondria



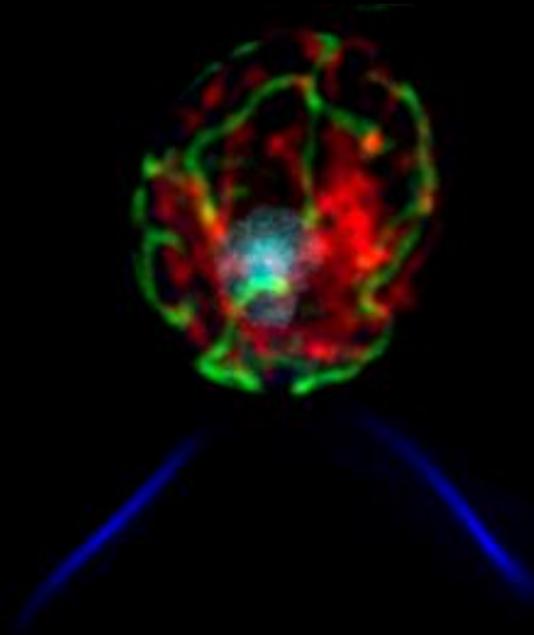
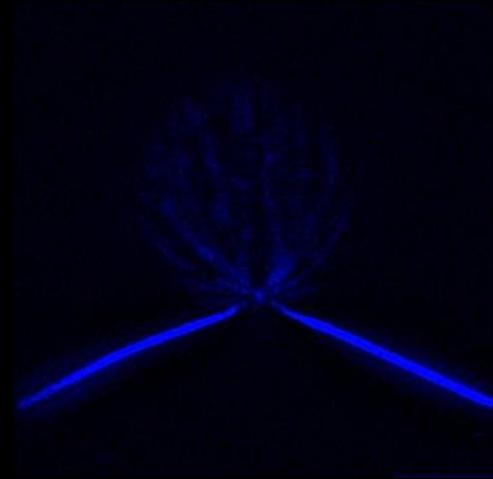
Gene stacking through mating



3rd
X

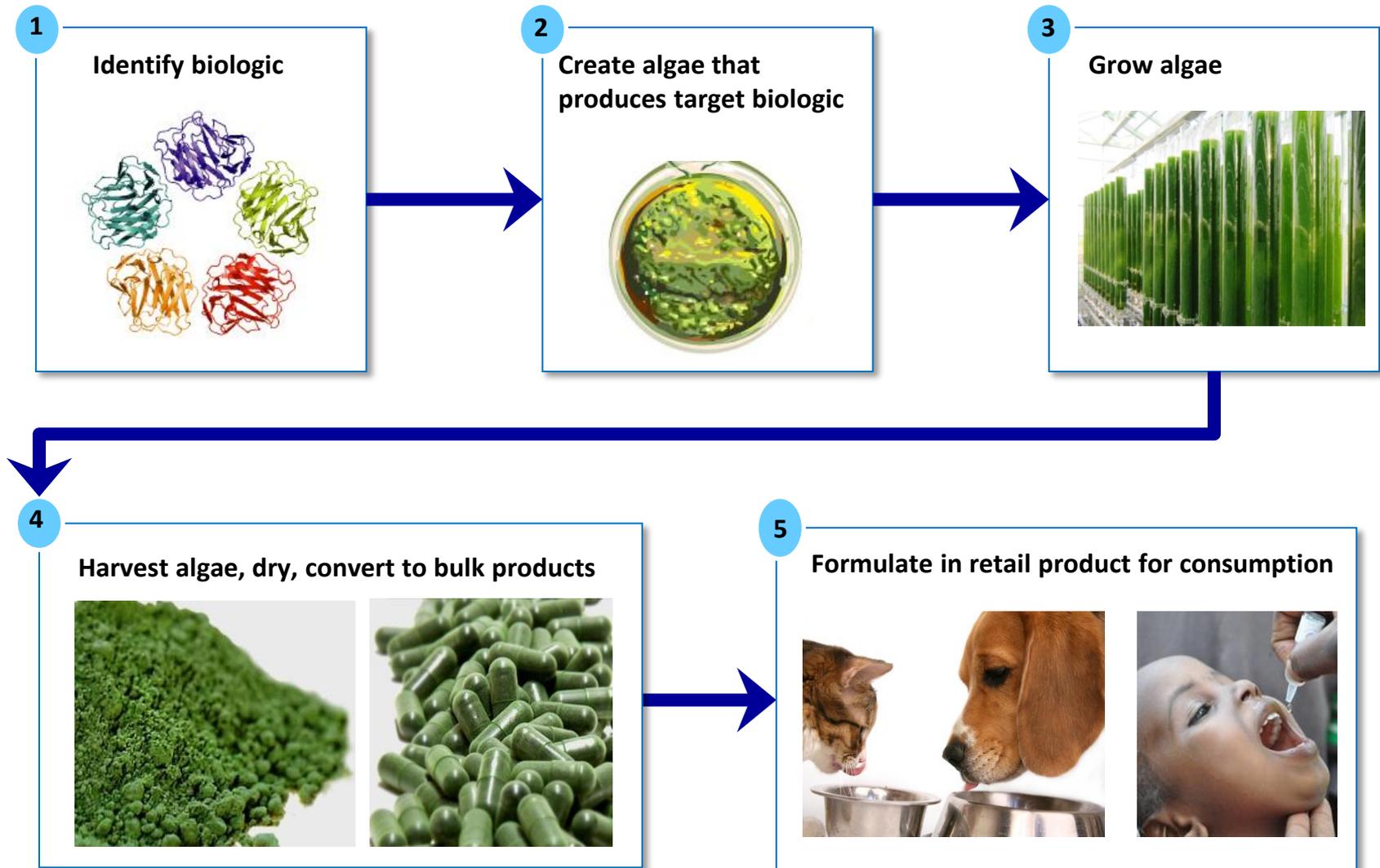


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- A natural product found in colostrum and milk, never before available at large scale
- Stimulates the production of mucin in the small intestines to block pathogenic bacterial infections
- Potential to replace the prophylactic use of antibiotics in livestock
- Produced at a cost that is affordable to people in the developing world
- Sales will begin in 2014, in partnership with



Acknowledgements

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The Mayfield Lab:

Prof. Stephen Mayfield
Gino Chao
Matthew Pier
Jenny Ng
Dan Barrera
Austin Hallgren

TRITON
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Jason Pyle, CEO
Stephen Mayfield
William Julien
Nick Hofmeister
Daniel Sachs
Miller Tran



Mike Mendez
Phil Lee



Todd Peterson
Farzad Haerizadeh
Lisa Stillwell



UC San Diego

Steve Briggs
Zhouxin Shen
Mark Hildebrand
Susan Golden
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Federico Unglaub

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